

INVE
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SURVEY on SHRIMP POST-LARVAL QUALITY:

ECUADORIAN versus OTHER LATIN-AMERICAN and SOUTH-EAST ASIAN hatcheries

E. Werbrouck¹, R. Wouters¹, E. Naessens¹, M. de Lourdes Cobo Barcia², T. García², V. Toledo², G. Chauvet³, J. J. Muñoz-Medina³, T. Santos Silva⁴, M. Santos⁴, C. Sankaranarayanan⁵, M. Rahman⁵, M. Janssens⁵, E. Van Ballaer⁵, Chien Ho Van⁵, B. A. Narlioglu⁵, P. Lavens¹

¹ INVE Technologies NV Belgium

² INVE del Ecuador Cdla

³ INVE Aquaculture Mexico S.A. De C.V.

⁴ INVE do Brasil Ltda.

⁵ INVE Asia Services Ltd.

Wednesday 25th October
10:00 – 10:50

1. Introduction

- 1.1. What is shrimp post-larval quality (=PL Q)?
- 1.2. Survey build-up and global distribution
- 1.3. Data analyses: Ecuador versus other Latin-American countries (=other LA) versus South-East Asia (=SEA)

2. Results

- 2.1. Hatcheries' characteristics
- 2.2. Opinions on shrimp PL Q
 - 2.2.1. Awareness
 - 2.2.2. Determining factors
 - 2.2.3. Long term effects
 - 2.2.4. Financial implications
 - 2.2.5. Disease incidence
- 2.3. PL Q parameters
 - 2.3.1. Monitoring frequency
 - 2.3.2. Relevance to estimate PL Q
 - 2.3.3. Relevance to predict grow-out (=GO) performance

3. Conclusions

4. Future

1. INTRODUCTION

1.1. What is shrimp PL Q?

the 'readiness' of post-larvae to be stocked in ponds

the physiological condition of shrimp post-larvae, their performance during culture, and resistance to stress tests
(Racotta et al 2003)



No well-defined
universally-accepted
evaluation method

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1. INTRODUCTION

1.1. What is shrimp PL Q?

'Inferior post-larvae have played a part in almost 80% of Early Mortality Syndrome occurrence in Malaysia'

(K. Muthusamy in AQUA Culture AsiaPacific, 2013)

'Production and profitability of shrimp farms can be increased by stocking only high quality post-larvae'

(D.E. Jory in Global Aquaculture Alliance, 2017)

'Transition of the hatchery stage from a secondary role into the starring role'

(McIntosh, CPF Thailand – Larvi Conference, 2017)



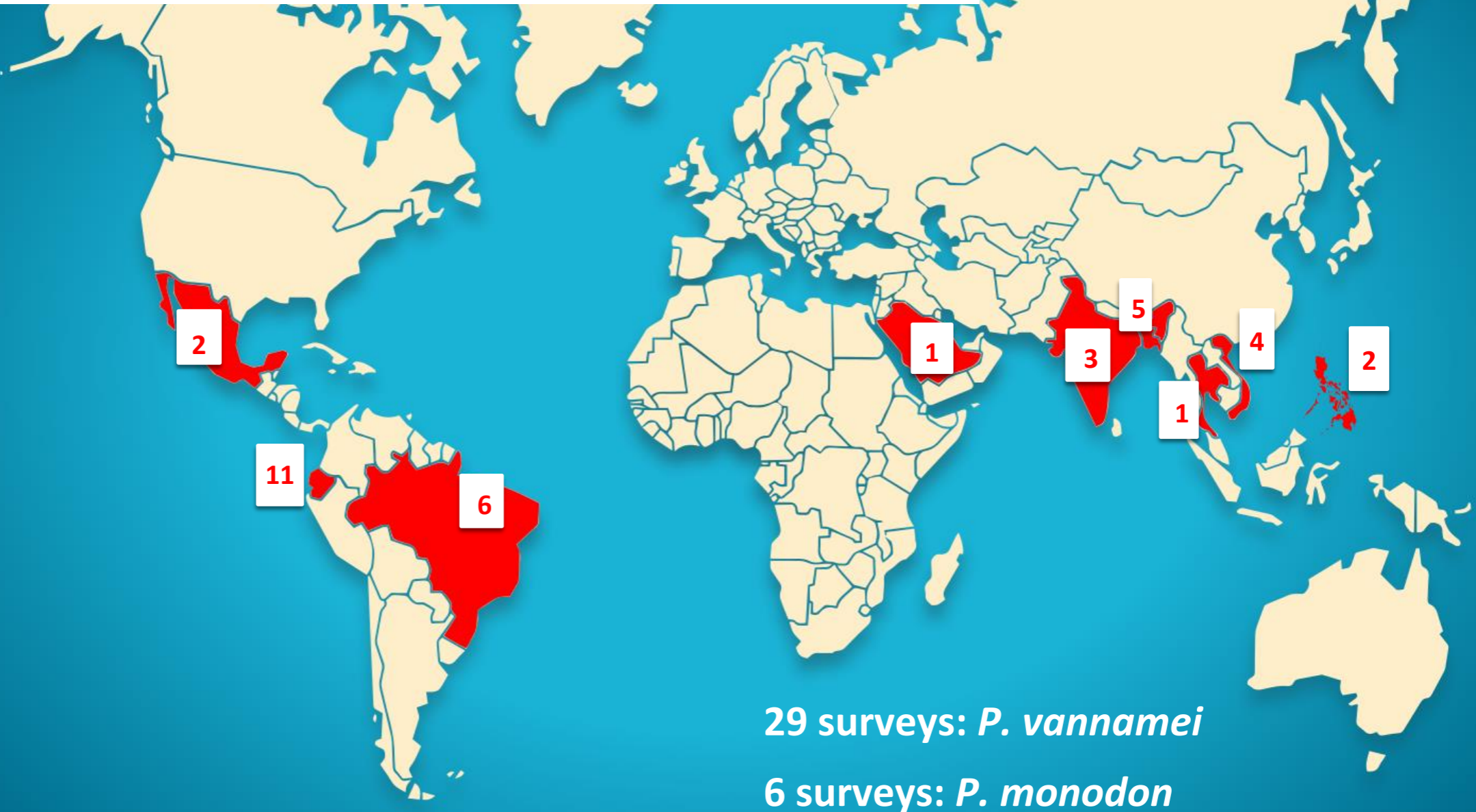
Survey on PL Q opinions and practices
in commercial hatcheries

1. INTRODUCTION

1.2. Survey build-up and global distribution (total 35 surveys)

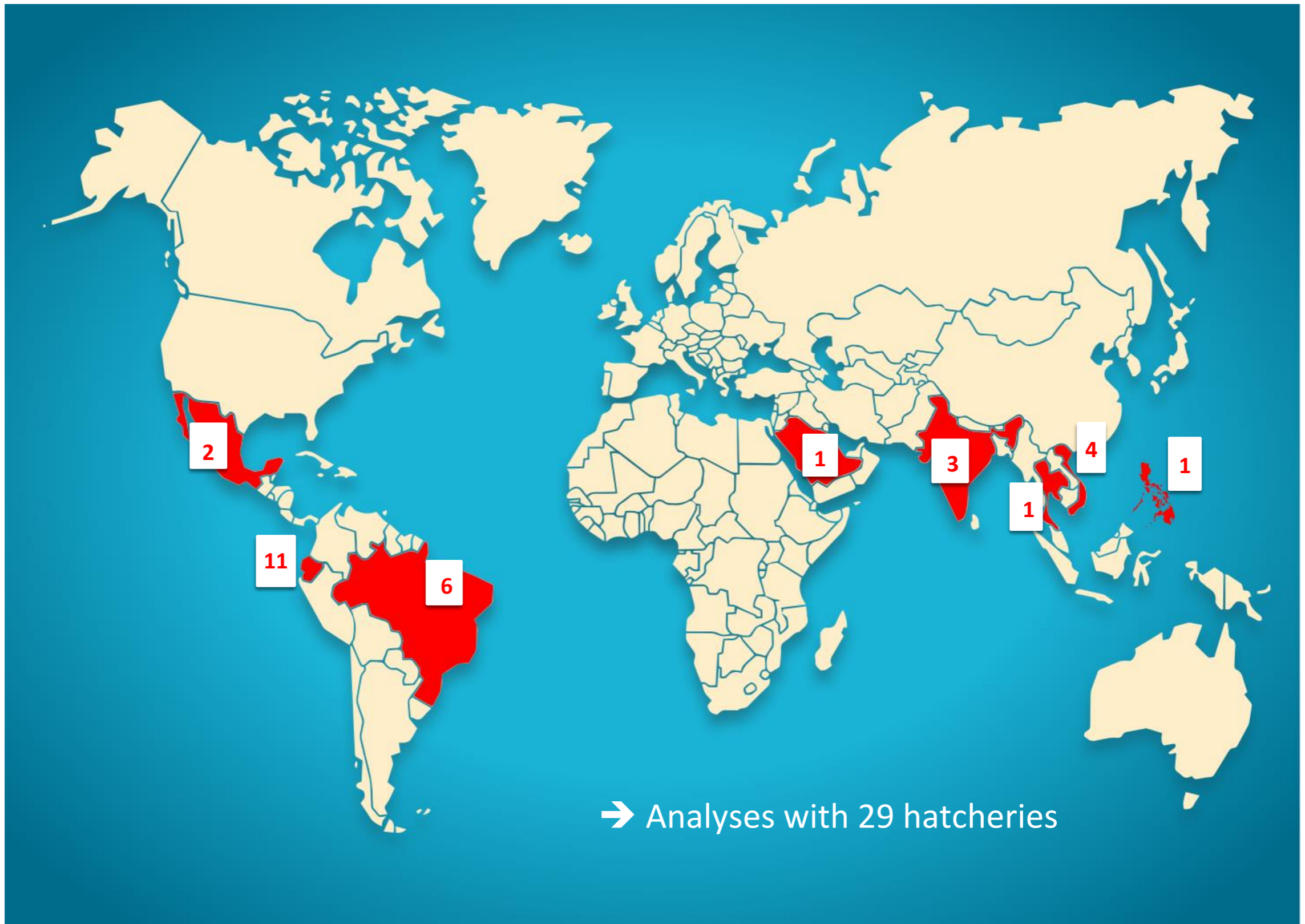
Focus: PL Q opinions and practices

Extended: technical questions on the culture practices



1. INTRODUCTION

1.2. Survey build-up and global distribution - *P. vannamei*



1. INTRODUCTION

1.3. Data analyses: Ecuador versus other LA versus SEA





2. RESULTS

2.1. Hatcheries' characteristics



2. RESULTS

2.1. Hatcheries' characteristics: Ecuador – other LA – SEA

stocking density

| | avg nps/ L | min - max range |
|----------|------------|-----------------|
| Ecuador | 144 | 90 - 200 |
| other LA | 276 | 150 - 400 |
| SEA | 187 | 100 - 300 |

survival at the end of the culture cycle

| | avg (%) | min - max range |
|----------|---------|-----------------|
| Ecuador | 63 | 55 - 75 |
| other LA | 59 | 45 - 75 |
| SEA | 46 | 30 - 75 |

number of days to complete a culture cycle

| | avg days | min - max range |
|----------|----------|-----------------|
| Ecuador | 22 | 19 - 25 |
| other LA | 21 | 18 - 23 |
| SEA | 20 | 18 - 25 |

own live algae production facility

| | avg (%) |
|----------|---------|
| Ecuador | 64 |
| other LA | 100 |
| SEA | 40 |

2-phase system in the hatchery phase

| | avg (%) |
|----------|---------|
| Ecuador | 55 |
| other LA | 88 |
| SEA | 50 |



2. RESULTS

2.1. Hatcheries' characteristics Ecuador – other LA – SEA

use of feed that color the hepatopancreas

| | avg (%) |
|----------|---------|
| Ecuador | 82 |
| other LA | 63 |
| SEA | 70 |

use of specialty feed with health claims*

| | avg (%) |
|----------|---------|
| Ecuador | 45 |
| other LA | 100 |
| SEA | 20 |

* High levels of vitamins, immunostimulants

sells only to 3rd party grow-out farms

| | avg (%) |
|----------|---------|
| Ecuador | 45 |
| other LA | 50 |
| SEA | 30 |



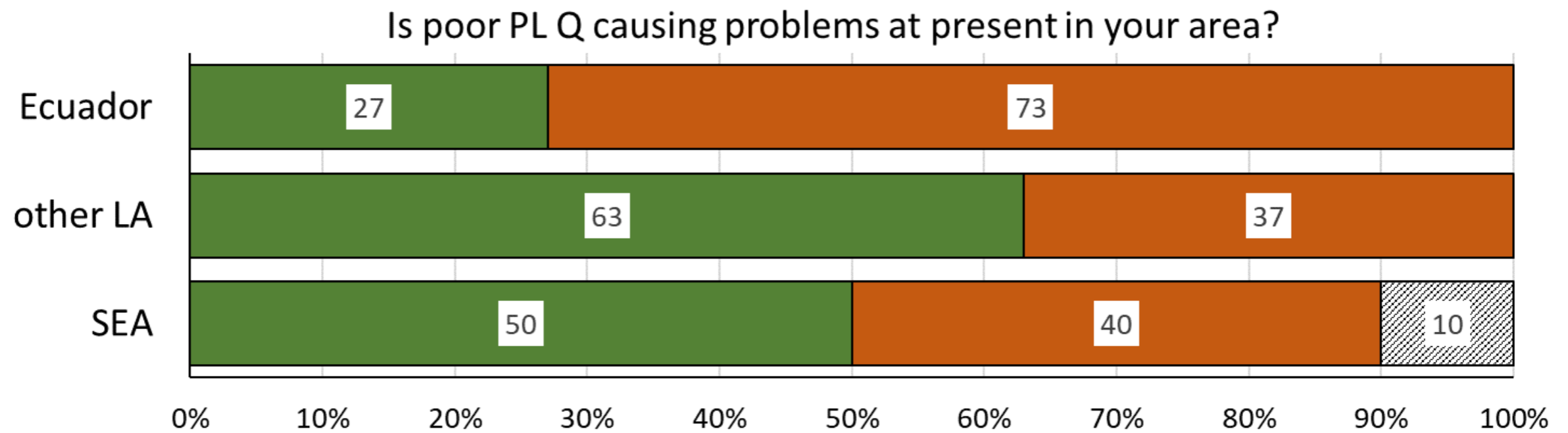
2. RESULTS

2.2. Opinions on shrimp PL Q



2.2.1. Awareness

■ yes ■ no ■ blank



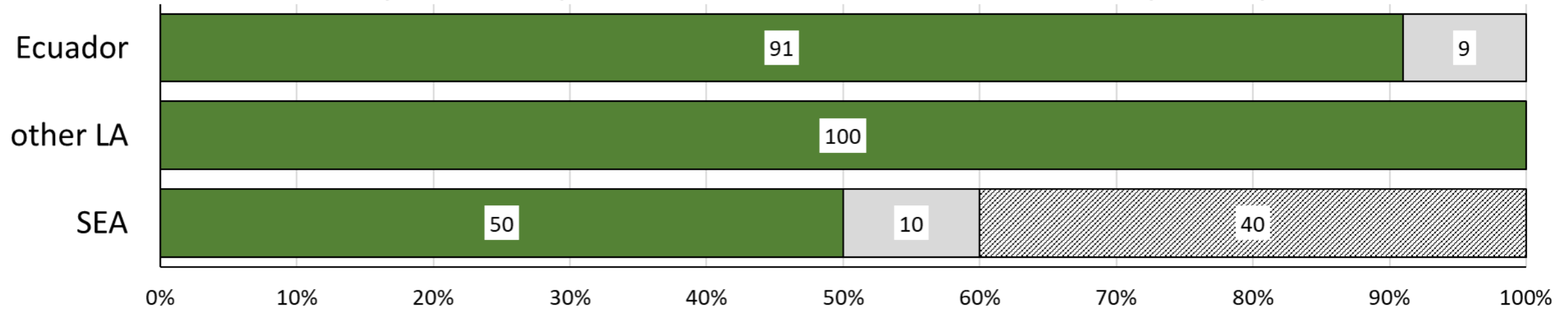
What % of screened batches is typically rejected due to PL Q problems?

| | avg (%) |
|----------|---------|
| Ecuador | 5 |
| other LA | 2 |
| SEA | 7 |

2.2.1. Awareness

■ improved
 ■ maintained
 ■ decreased
 blank

Has PL Q in your area improved, maintained or decreased over the past 10 years?

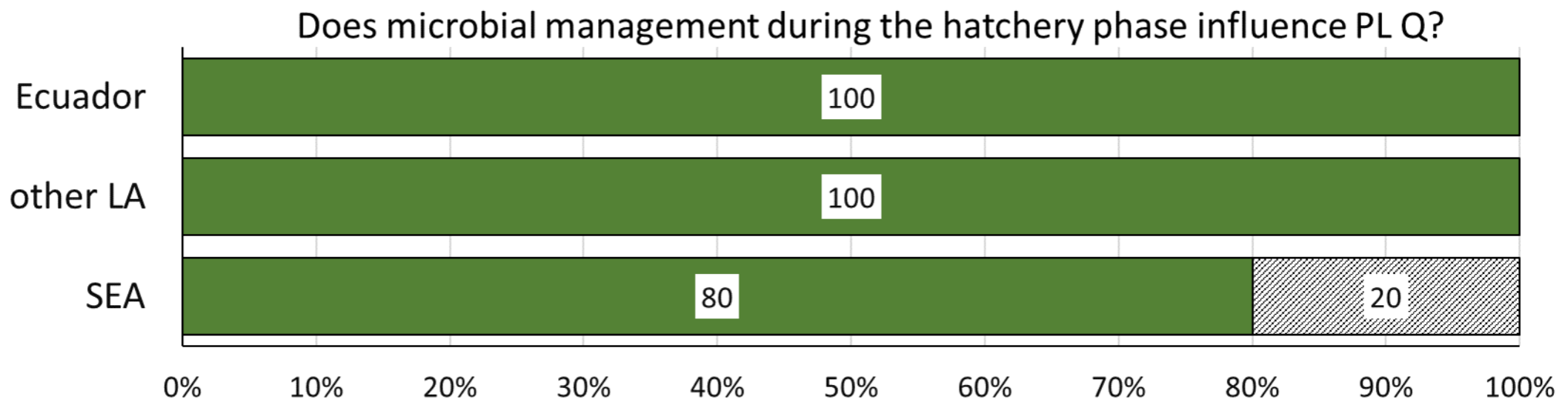
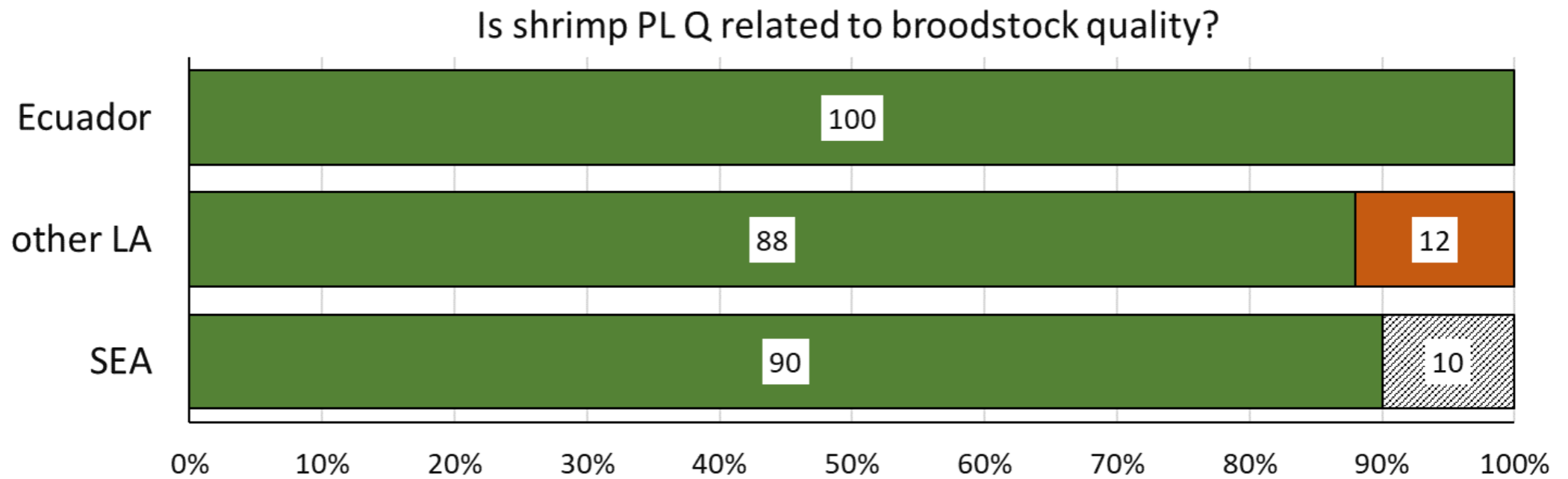


What % of the PL dies shortly after transfer from hatchery/ nursery to ponds?

| | avg (%) | min - max range |
|----------|---------|-----------------|
| Ecuador | 11 | 5 - 20 |
| other LA | 3 | 0 - 10 |
| SEA | 7 | 3 - 10 |

2.2.2. Determining factors

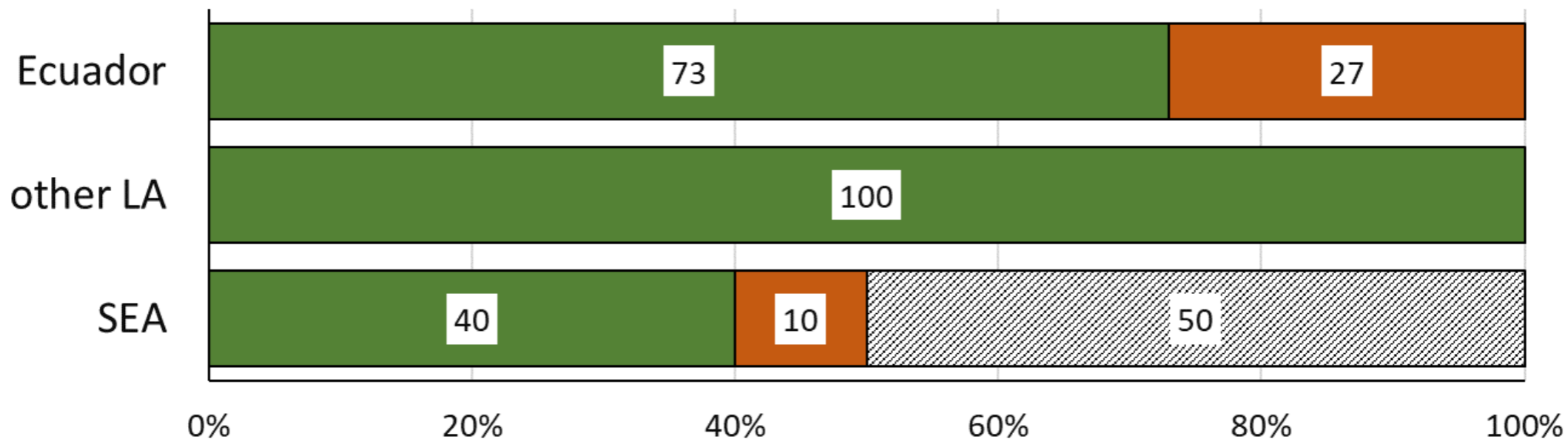
■ yes ■ no ■ blank



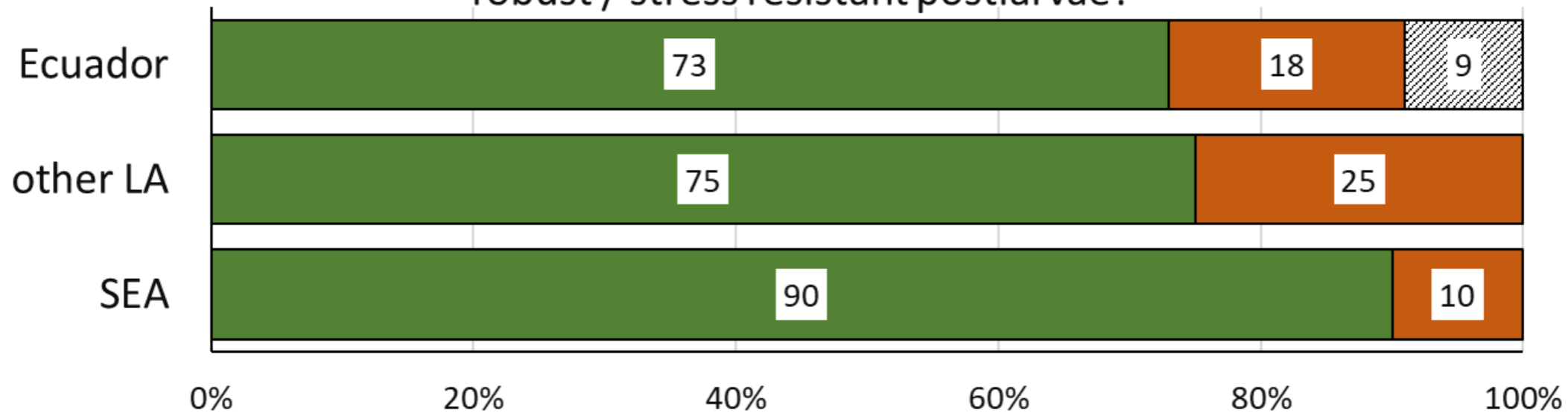
2.2.2. Determining factors

■ yes ■ no ■ blank

Do feeds with health claims produce more robust postlarvae?



Do you think that additives to control Vibrio levels produce more robust / stress resistant postlarvae?



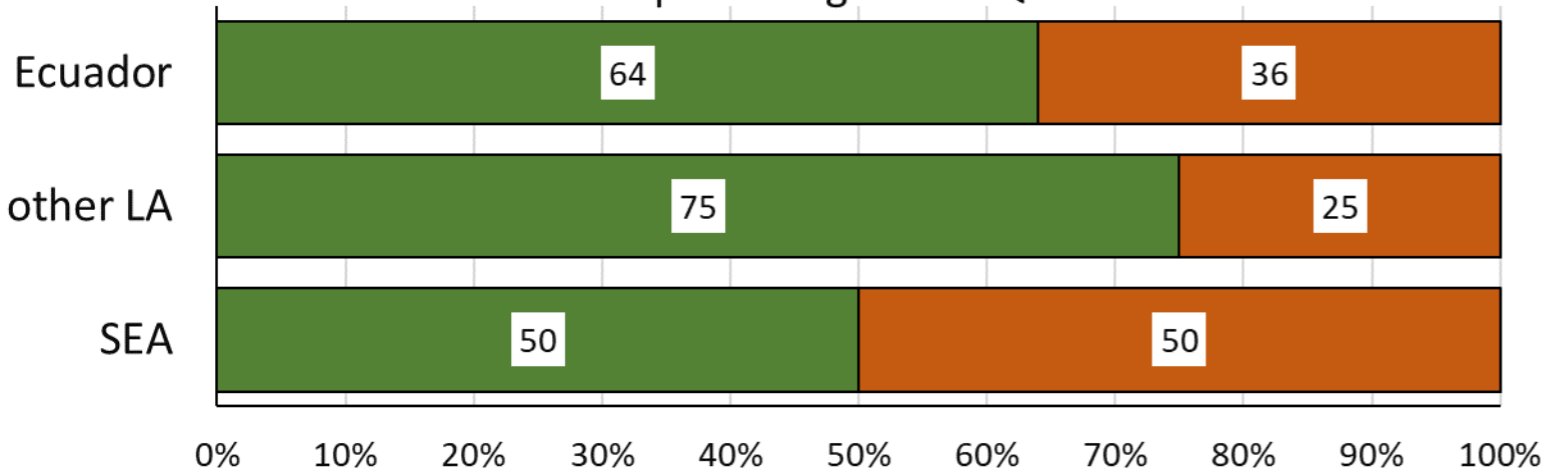


2.2.2. Determining factors

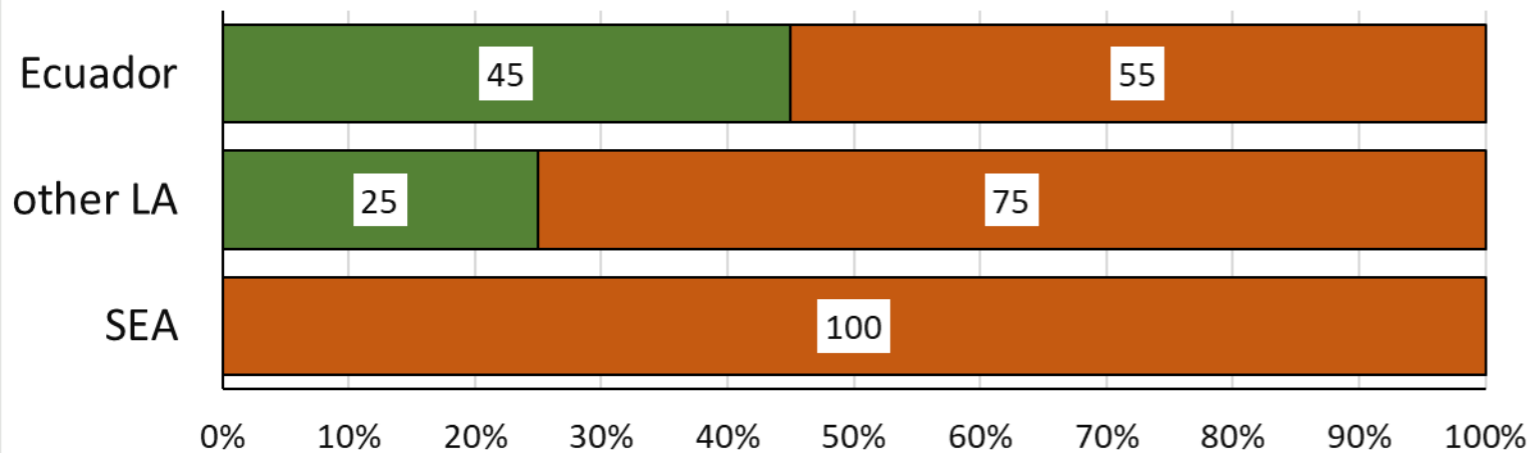
Do you think that ...

■ yes ■ no ■ blank

... feeding regimes with **REDUCED** Artemia consumption can produce good PL Q?



... feeding regimes **WITHOUT** Artemia can produce good PL Q?



| smallest tanks % | rectangular | |
|---------------------|-------------|------------|
| | flat bottom | U/V shaped |
| Ecuador | - | 91 |
| other LA | 13 | 75 |
| SEA | 40 | 30 |

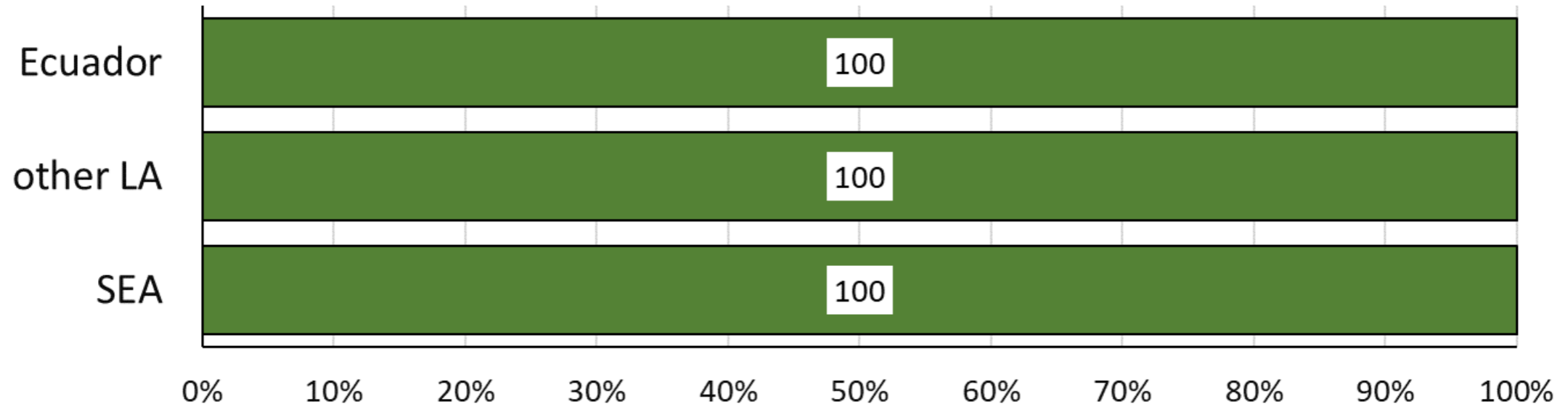
| largest tanks % | rectangular | |
|--------------------|-------------|------------|
| | flat bottom | U/V shaped |
| Ecuador | 18 | 64 |
| other LA | 25 | 38 |
| SEA | 40 | - |

2.2.3. Long term effects

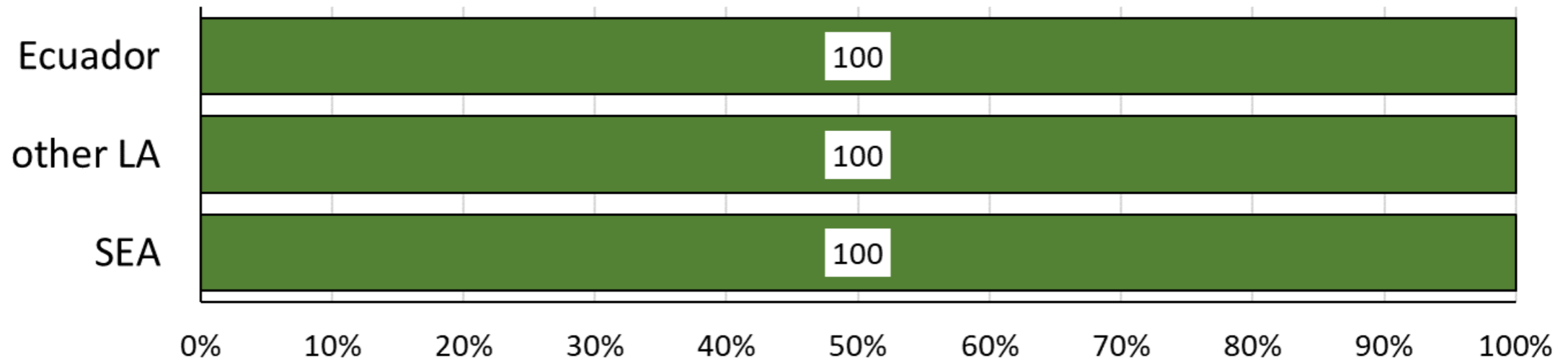
Do you think that ...

■ yes ■ no ■ blank

... differences in **PL Q** will reflect in the subsequent **nursery** phase?



... differences in **PL Q** will reflect in the subsequent **GO** phase?

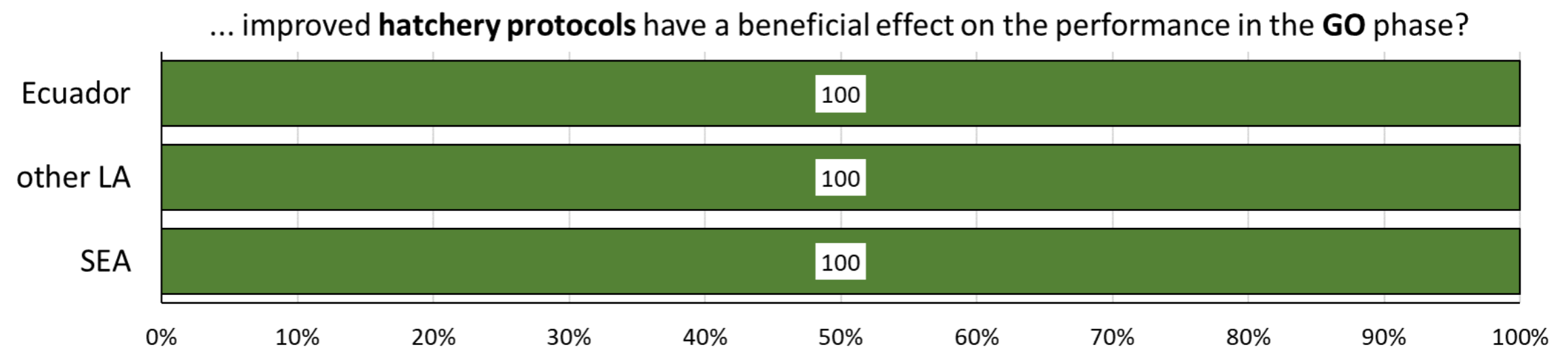
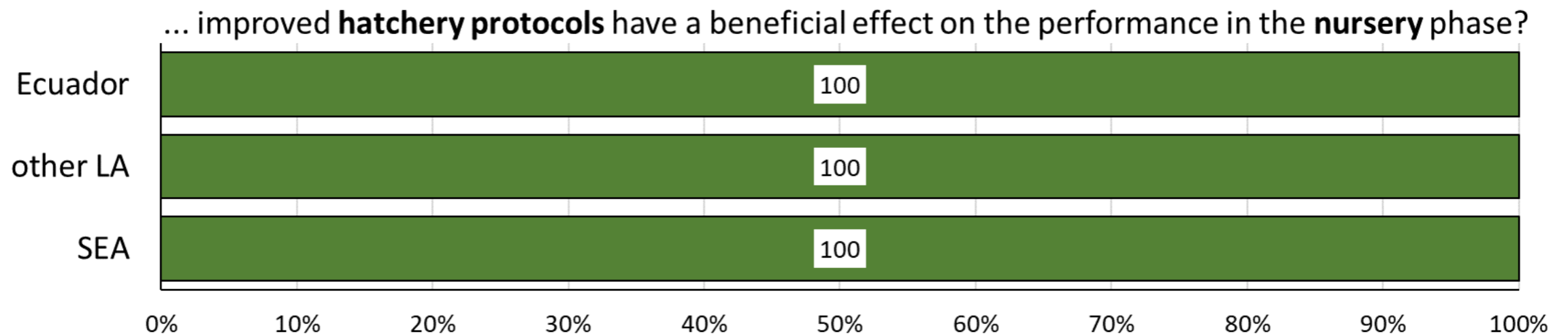




2.2.3. Long term effects

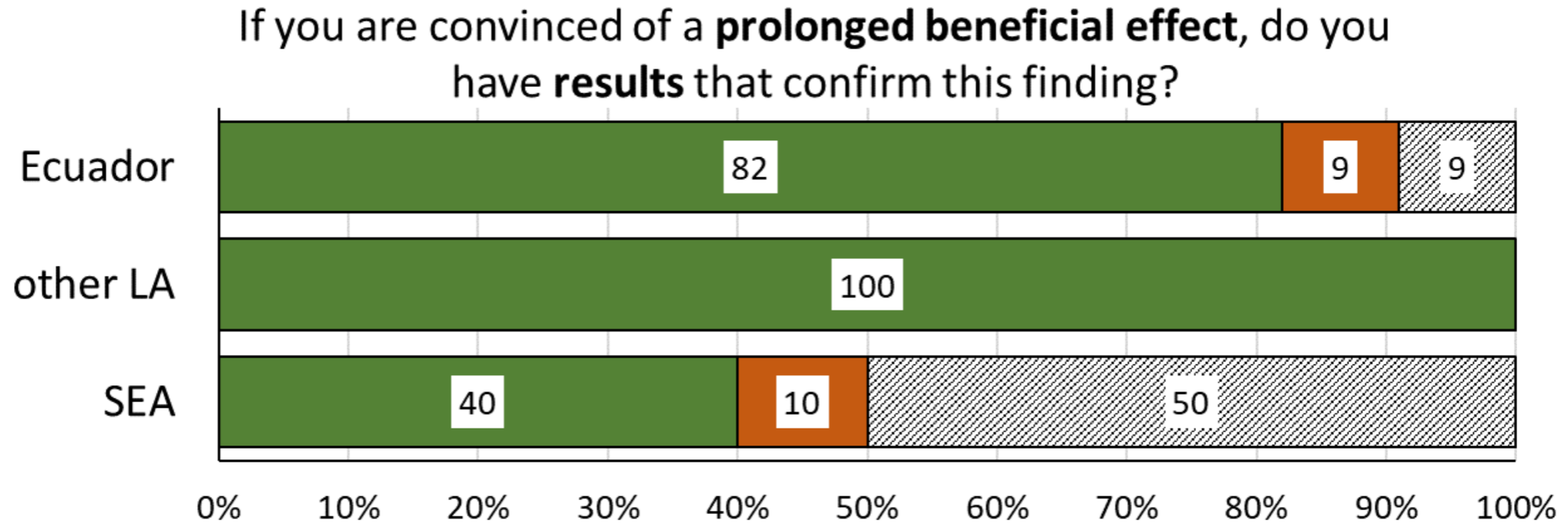
Do you think that ...

■ yes ■ no ■ blank



2.2.3. Long term effects

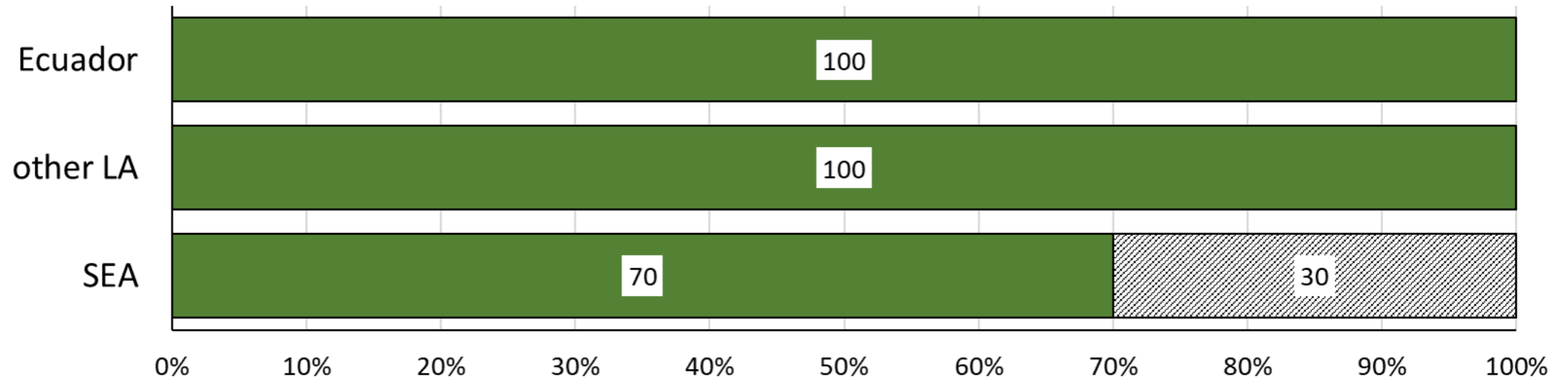
■ yes ■ no ■ blank



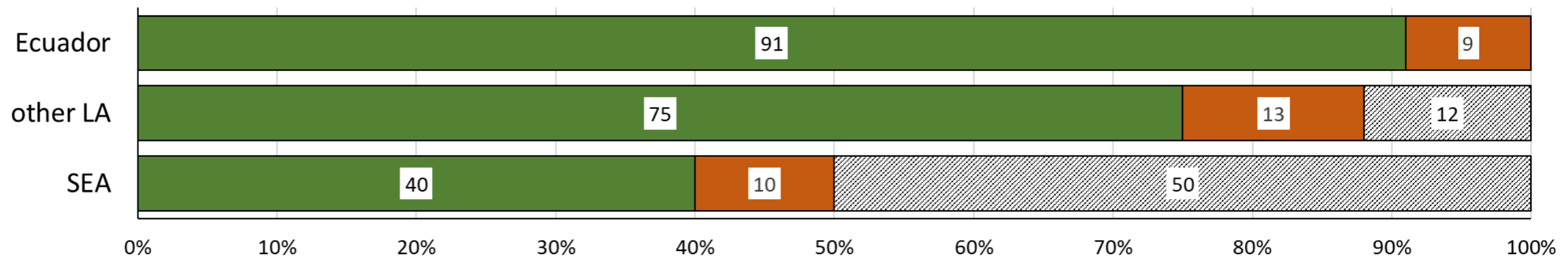
2.2.3. Long term effects

■ yes ■ no ■ blank

Can hatchery nutrition have a prolonged beneficial effect after stocking in ponds?



Can treatment of post-larvae to reduce Vibrio load have a prolonged beneficial effect after stocking in ponds?



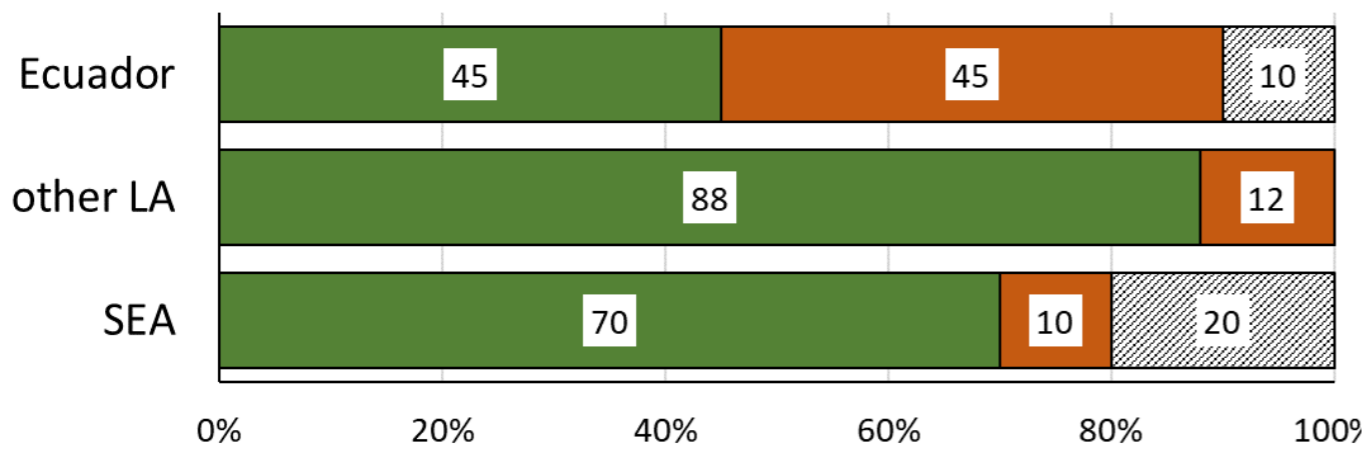


2.2.3. Long term effects

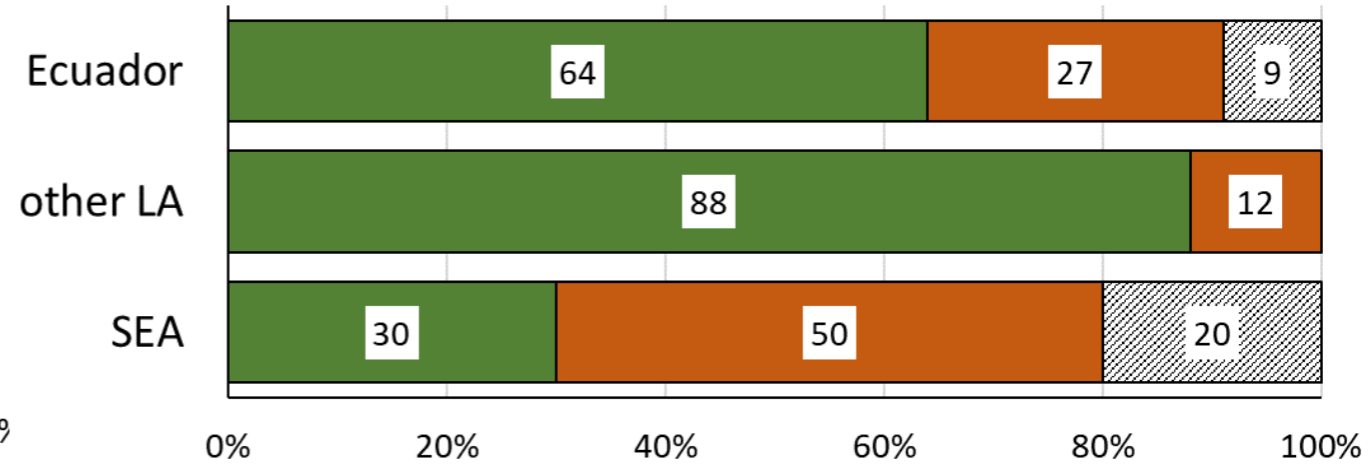
Which parameter do you think will be most affected by hatchery protocols:

■ yes ■ no ■ blank

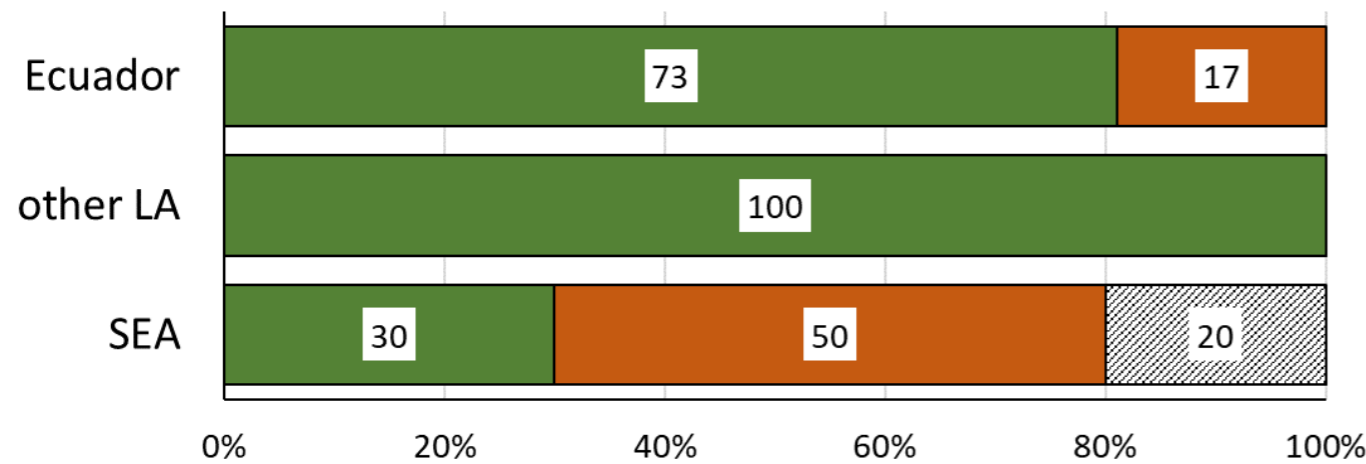
Disease resistance?



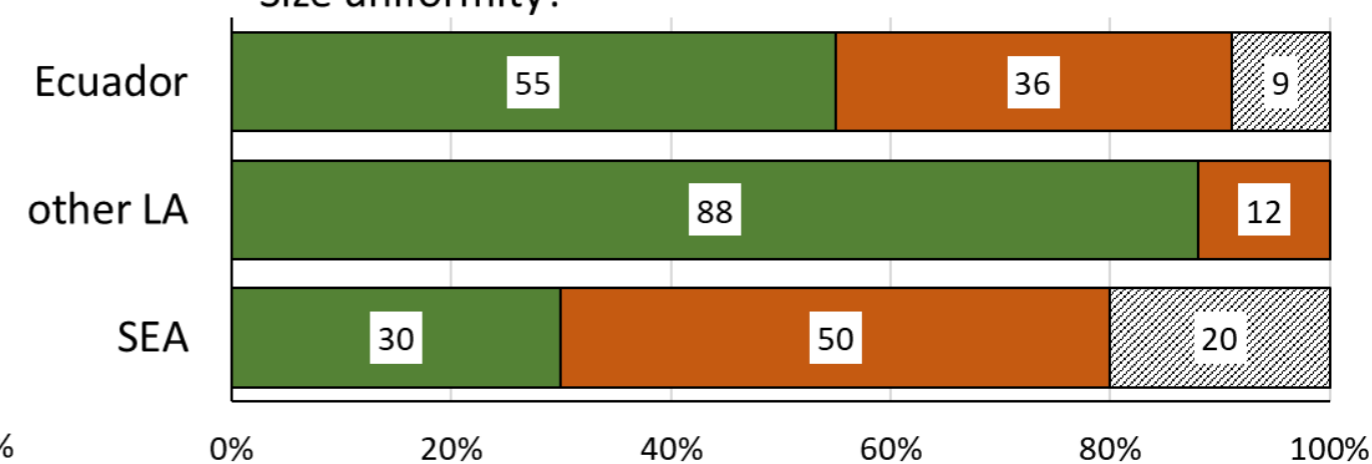
Survival?



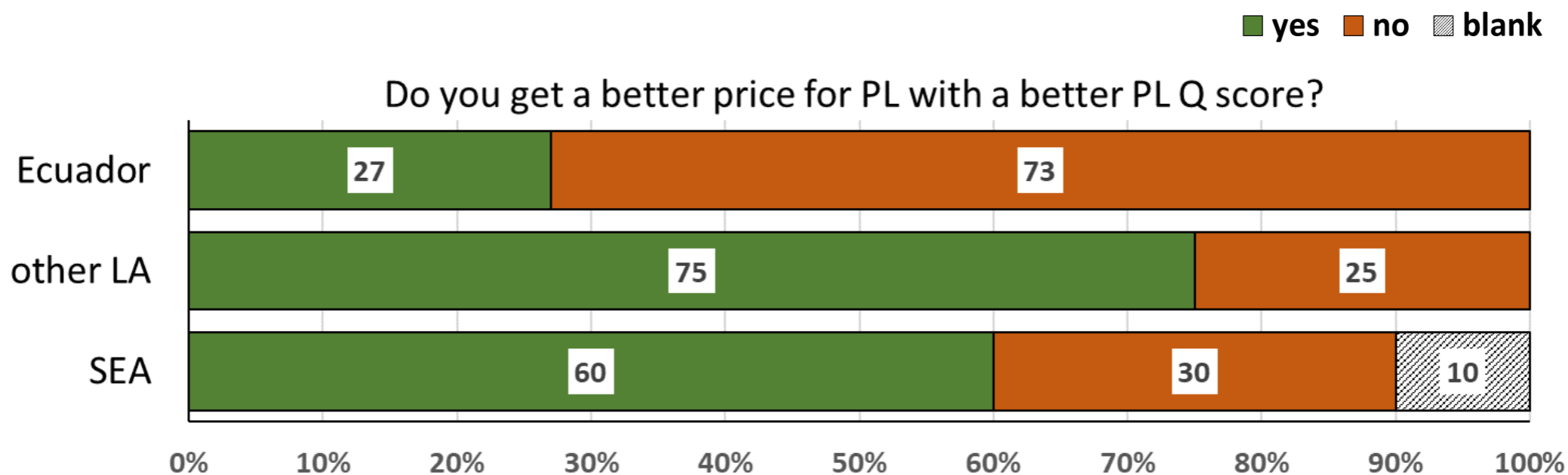
Growth?



Size uniformity?

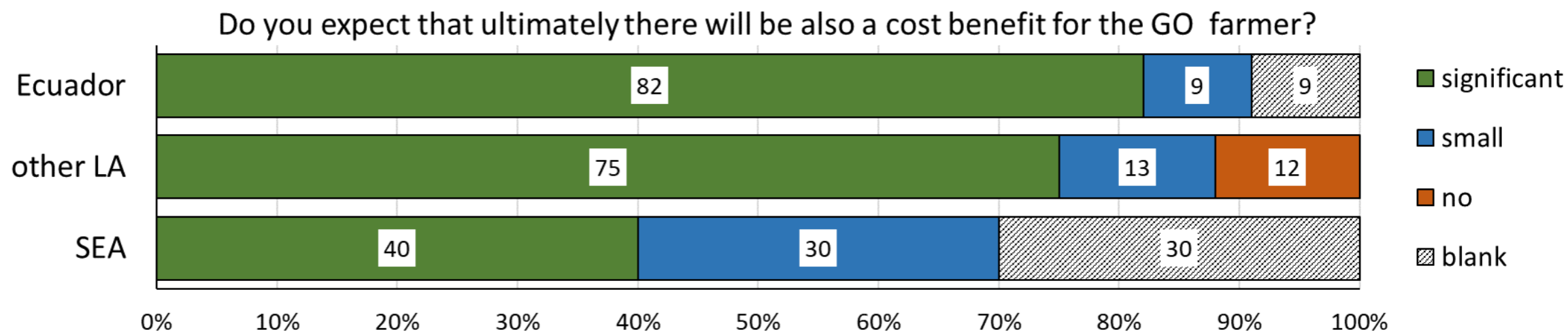


2.2.4. Financial implications

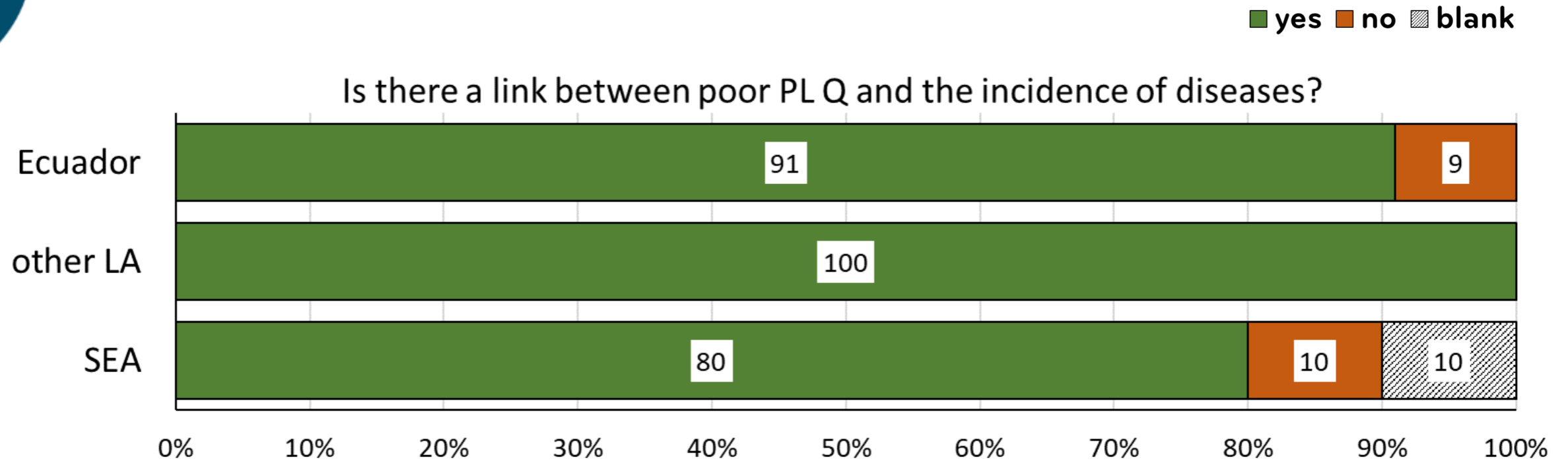


If yes, can you give an indication of how much the price gets (increase in %)?

| | avg (%) | min-max range |
|----------|---------|---------------|
| Ecuador | 10 | 5 - 15 |
| other LA | 19 | 4 - 41 |
| SEA | 15 | 5 - 30 |



2.2.5. Disease incidence



What % of the screened batches is typically rejected due to disease problems?

| | avg (%) | min - max range |
|----------|---------|-----------------|
| Ecuador | 3 | 0 - 10 |
| other LA | 2 | 0 - 10 |
| SEA | 9 | 0 - 20 |



2. RESULTS

2.3. PL Q parameters

2.3. PL Q parameters (29)

Hatchery

survival
larval development speed
PL count per gram wet weight
PL size (length)
PL stage (age)
nutritional history

Microscopical

lipid globules in HP
gill ramification
gut fulness
rostral counts
deformities, necrosis, fouling
molting (recent or not)
gut: muscle ratio
uropods
chromatophores

Visual

color HP
animal color
size variation (by eye)

(stress) tests

osmotic stress test
toxicity stress test
bacterial challenge test
counter current test

Behavioral

swimming activity
feeding activity

Analysis

total plate count (marine agar)
Vibrio count (TCBS or other)
screening for a pathogenic virus
biochemical analyses (lipid, glucose)

Index

multifactorial assessment

2.3. PL Q parameters

1) How frequent do you monitor ...?

never – occasionally – always

→ avg monitoring score

2) How important is the parameter to estimate PL Q?

not important – low – moderate – highest importance

→ avg score for estimating PL Q

3) How important is this parameter to predict performance in GO?

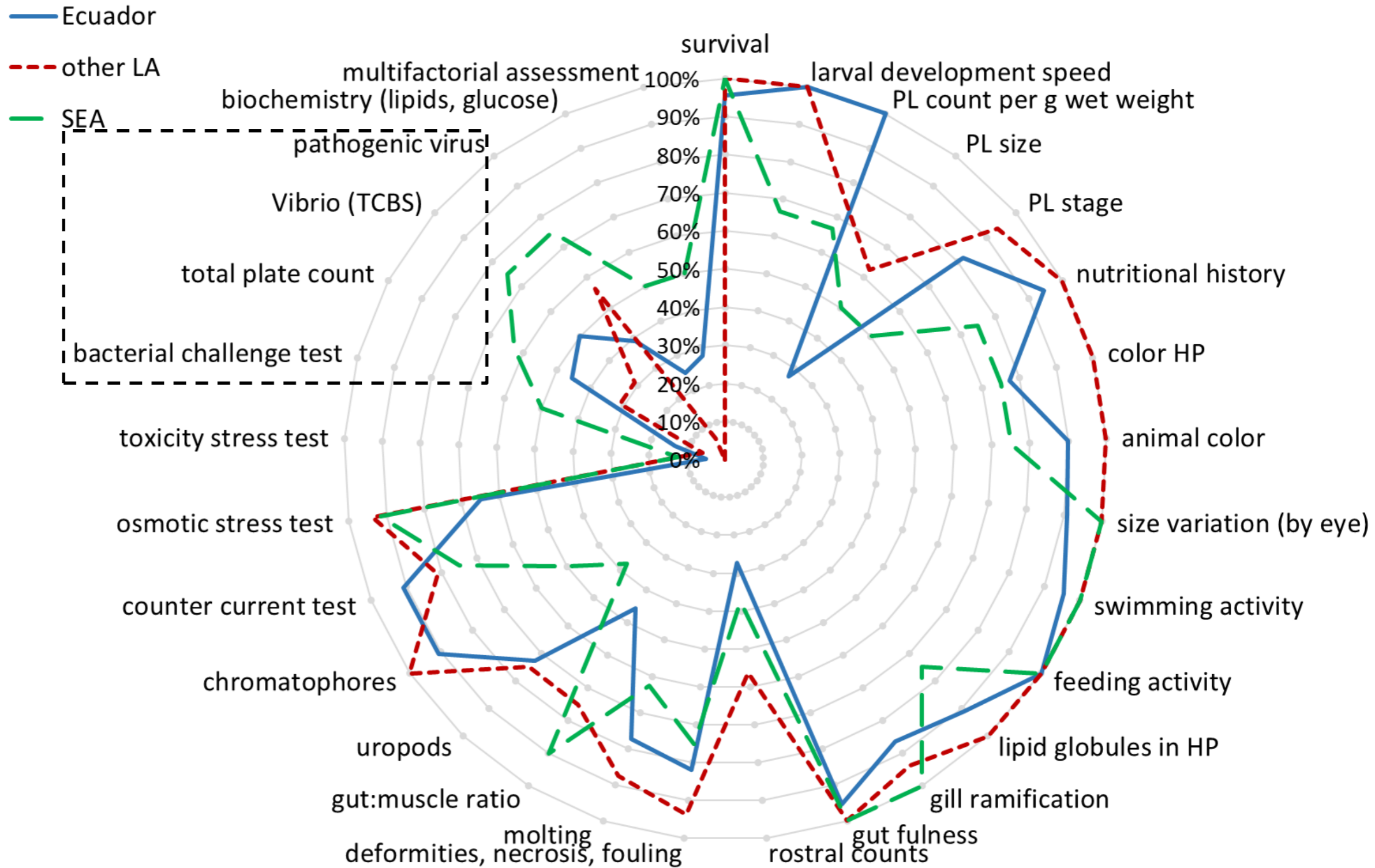
not important – low – moderate – highest importance

→ avg score for predicting GO performance

2.3.1. MONITORING – importance PL Q – importance GO performance

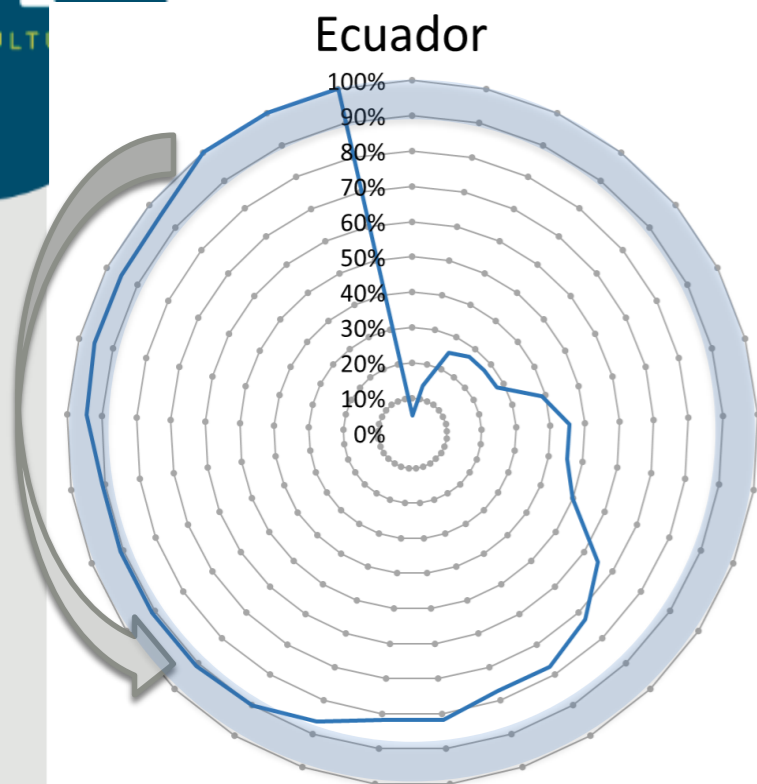


How frequently do you monitor ... ?

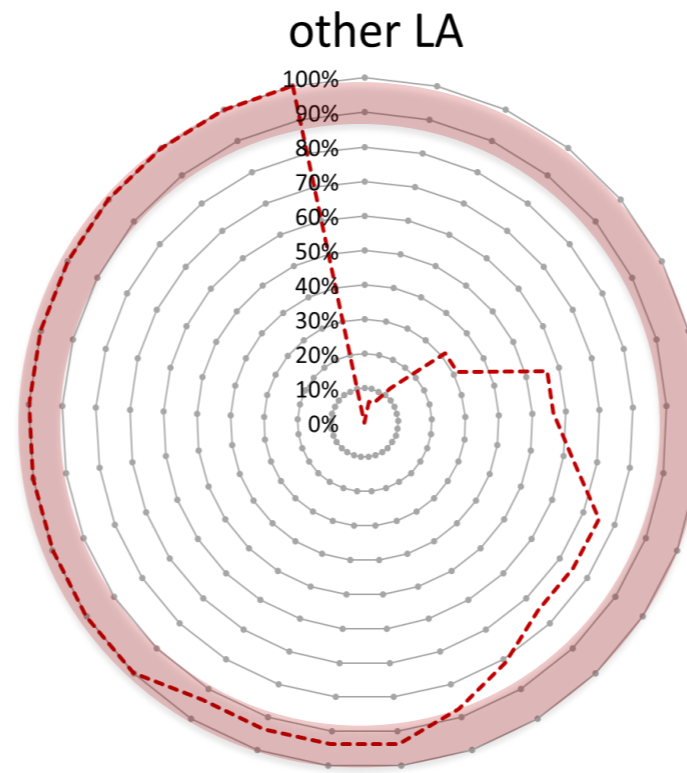


2.3.1. MONITORING – importance PL Q – importance GO performance

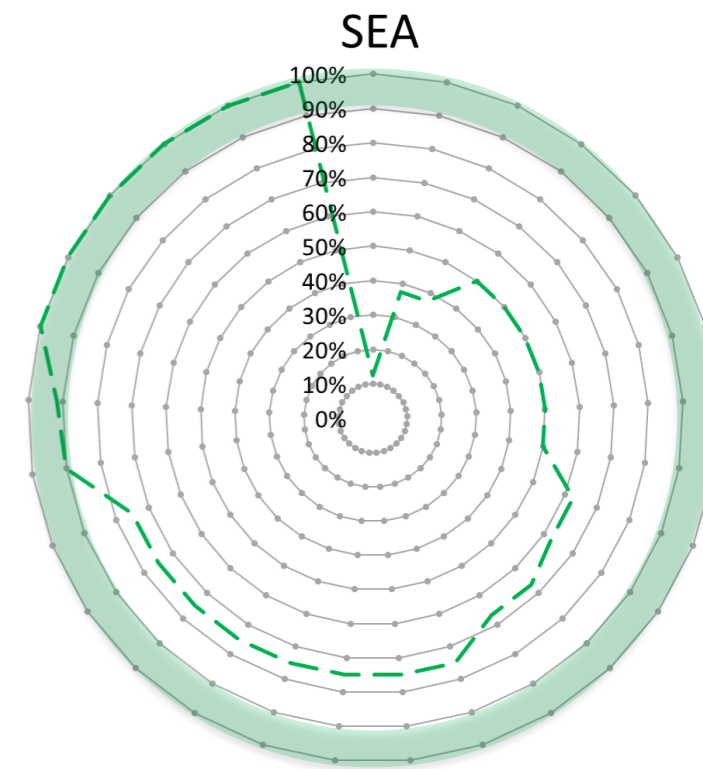
➔ most frequently monitored parameters (score $\geq 90\%$)



counter current test
PL count per g wet weight



PL stage
deformities, necrosis, fouling
color of HP



gut:muscle ratio

lipid globules in HP
nutritional history
larval development
chromatophores
animal color

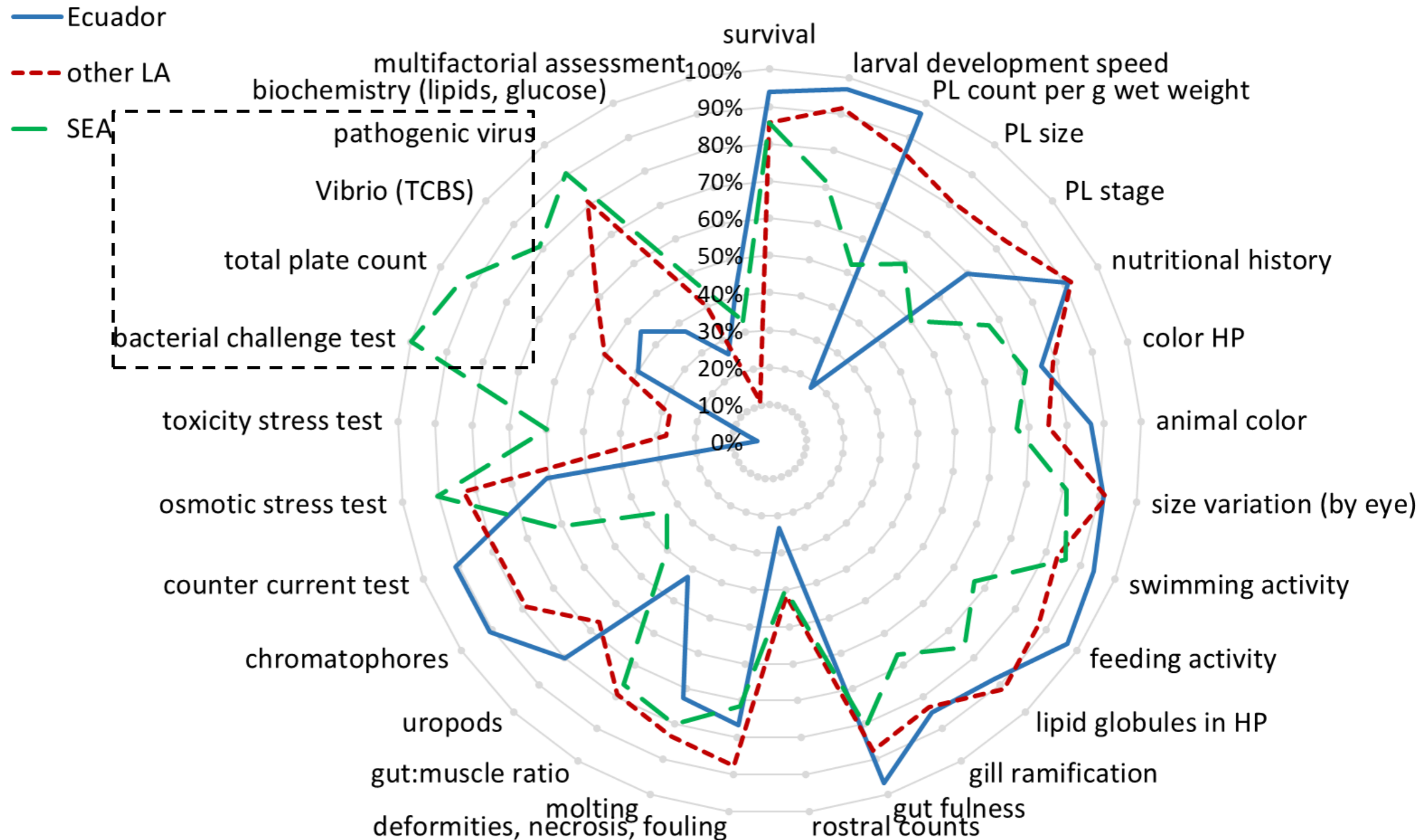
osmotic stress test
gill ramification

survival
size variation (by eye)
swimming activity
gut fulness
feeding activity

2.3.2. monitoring – IMPORTANCE PL Q – importance GO performance

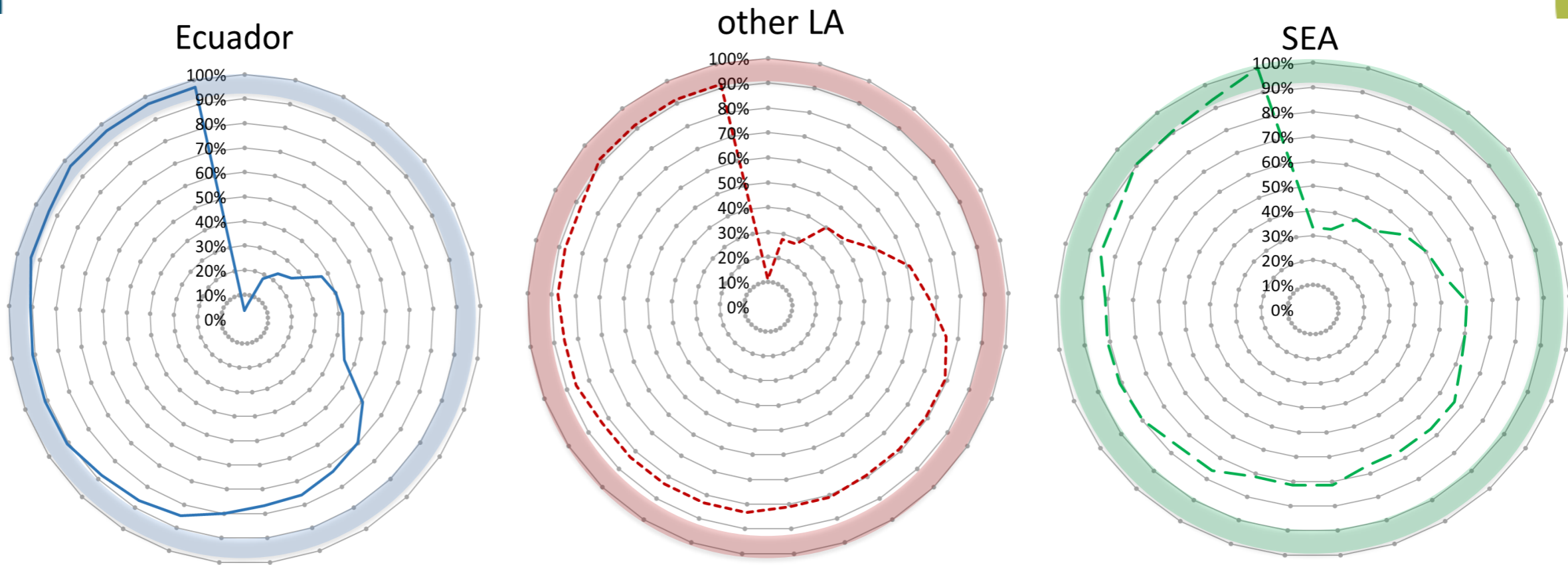


How important is the parameter to estimate PL Q?



2.3.2. monitoring – **IMPORTANCE PL Q** – importance GO performance

➔ most important parameters to estimate PL Q (score $\geq 90\%$)



| |
|---------------------------|
| chromatophores |
| counter current test |
| survival |
| swimming activity |
| feeding activity |
| gut fulness |
| PL count per g wet weight |

lipid globules in HP

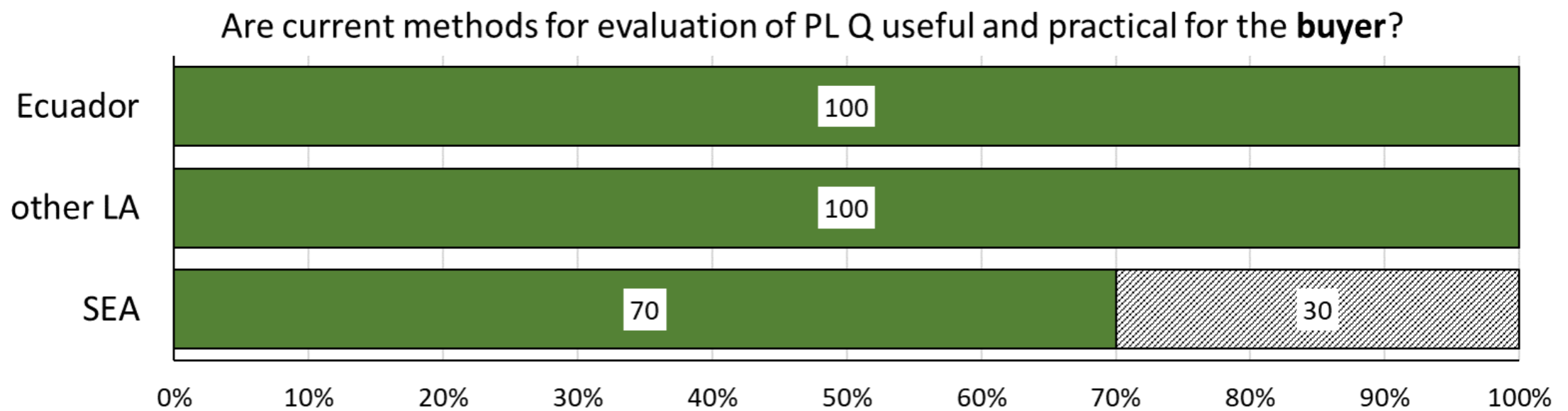
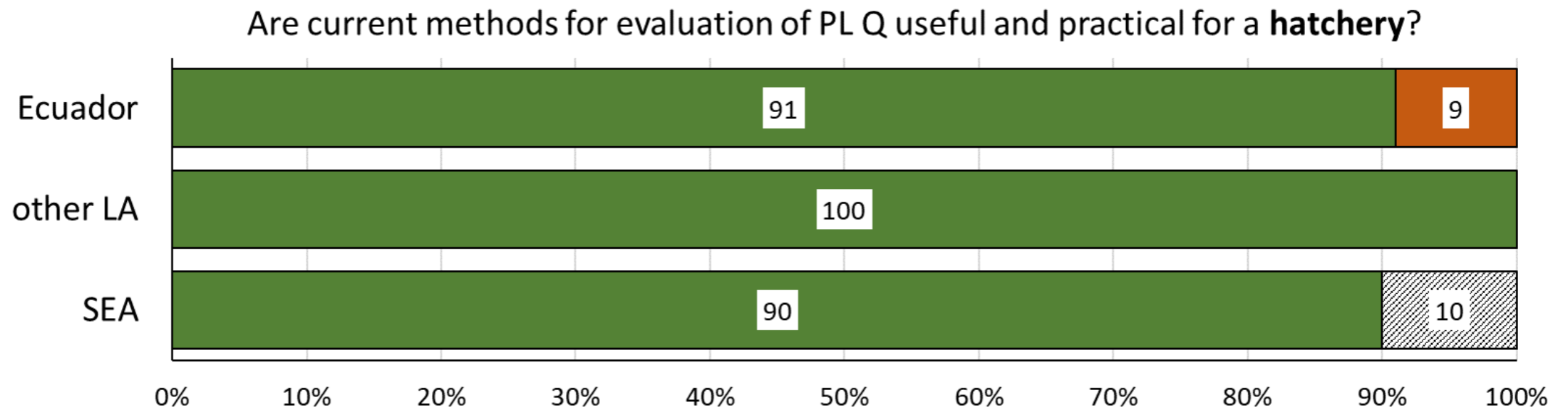
| |
|---------------------------------|
| osmotic stress test |
| pathogenic virus |
| total plate count (marine agar) |
| bacterial challenge test |

| |
|--------------------------|
| larval development speed |
| nutritional history |
| size variation (by eye) |

2.3.2. monitoring – **IMPORTANCE PL Q** – importance GO performance



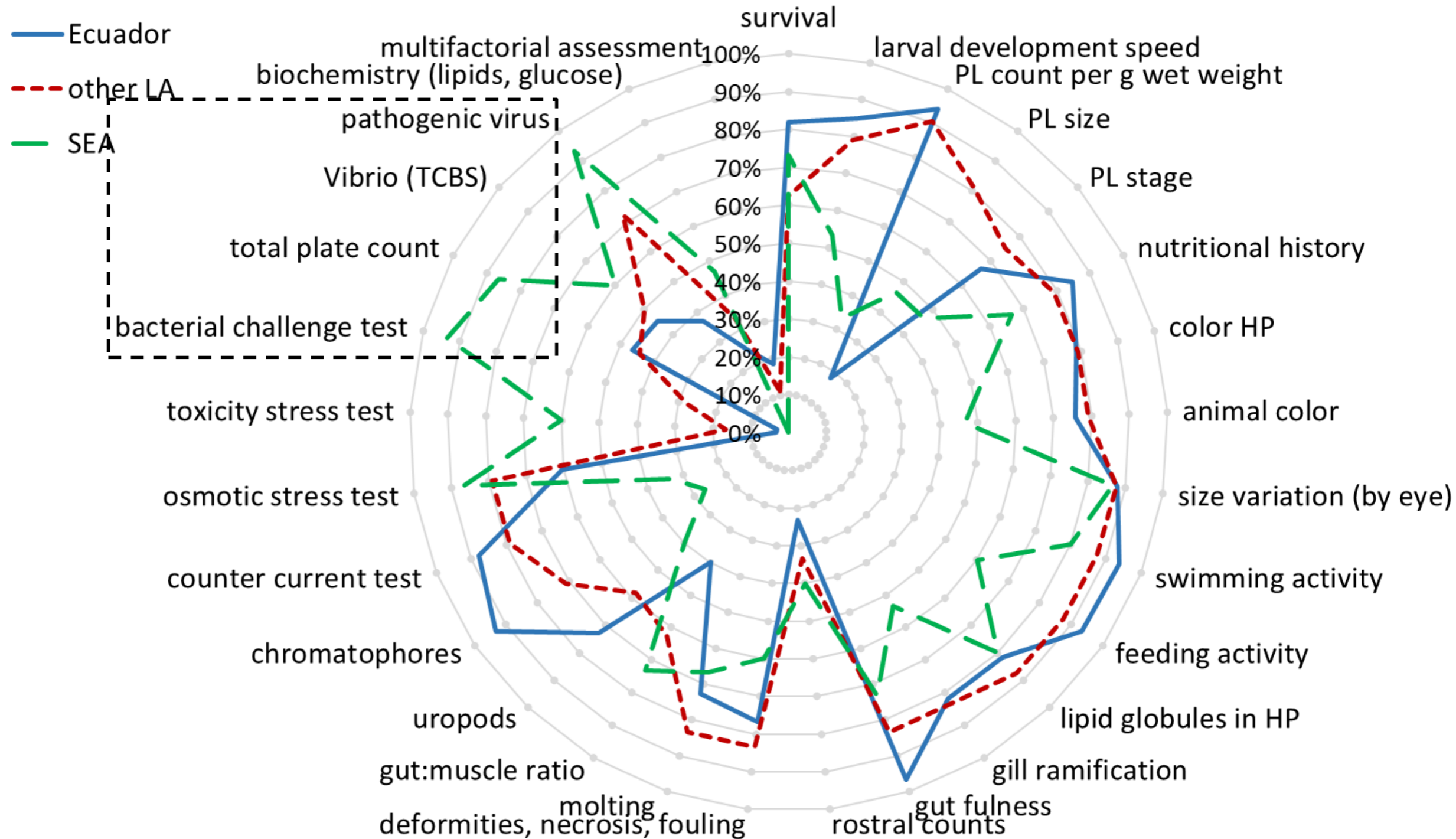
■ yes ■ no ■ blank



2.3.3. monitoring – importance PL Q – IMPORTANCE GO PERFORMANCE

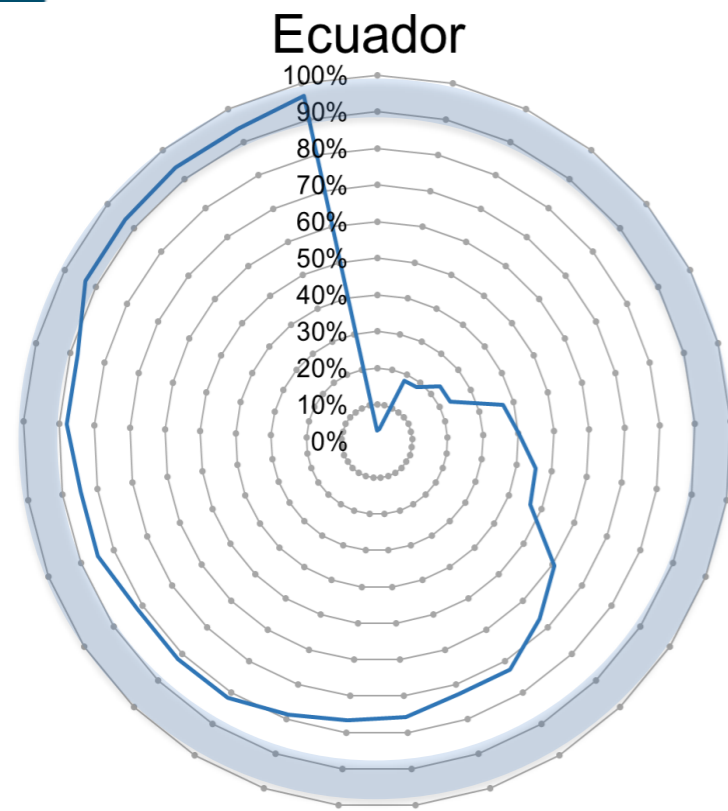


How important is the parameter to predict GO performance?

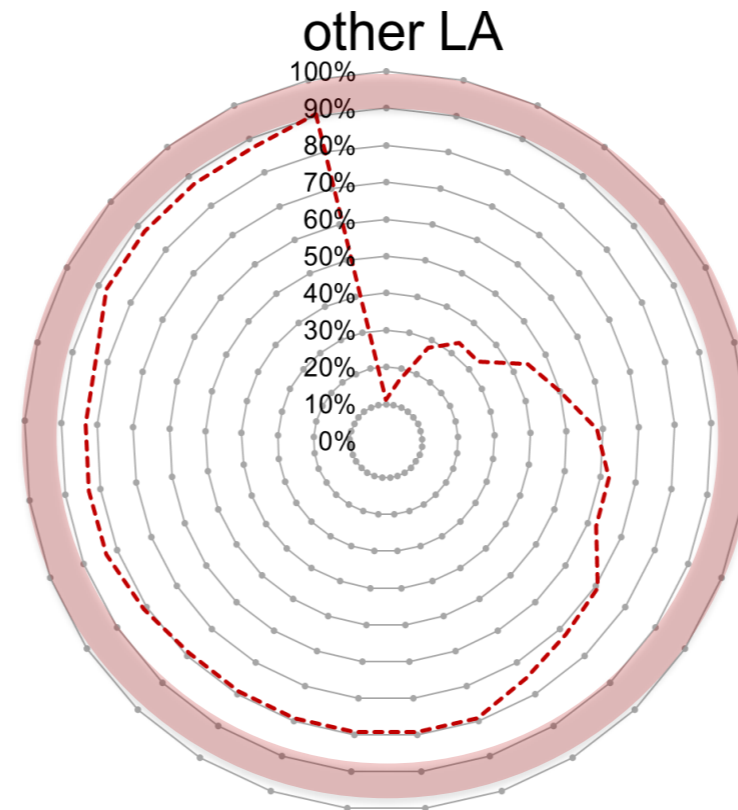


2.3.3. monitoring – importance PL Q – **IMPORTANCE GO PERFORMANCE**

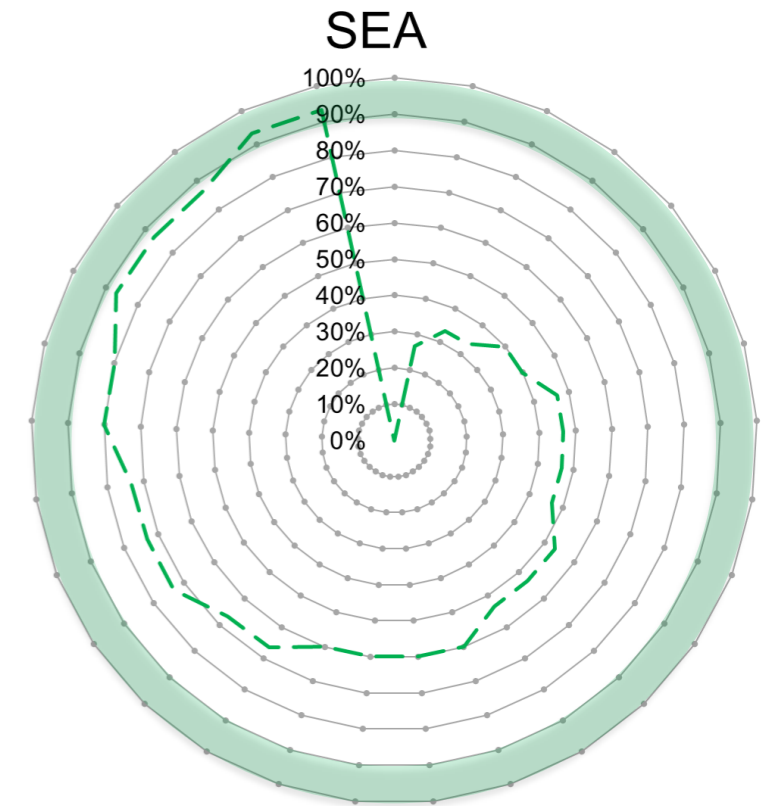
➔ **most important parameters for predicting GO performance (score ≥ 90%)**



| |
|-------------------|
| feeding activity |
| chromatophores |
| swimming activity |
| gut fulness |



| | |
|-------------------------|-----|
| swimming activity | 88% |
| feeding activity | 88% |
| size variation (by eye) | 88% |
| lipid globules in HP | 88% |



| |
|--------------------------|
| bacterial challenge test |
| pathogenic virus |

PL count per g wet weight

3. Conclusions

- high awareness on concept PL Q and its importance for predicting GO performance
- frequently monitored parameters worldwide:
survival, size variation (by eye), feeding and swimming activity, and gut fullness

little consensus on the most important parameters to estimate PL Q and to predict GO performance:

- ➔ higher importance of microbial parameters in SEA: site-specific challenges?
- ➔ interpretation PL Q may evolve over time?

3. Conclusions

Support for the following claims:

- strong link between PL quality and the incidence of disease
- differences in PL quality are reflected in nursery and grow-out phase
- improved hatchery protocols have a beneficial effect in nursery and grow-out phase
 - ➔ The majority in Latin-America (including Ecuador) has results that confirm this beneficial effect
- PLs of higher quality imply a cost-benefit for the grow-out farmer

3. Conclusions

Cost to produce 1 million PL → 15.5 ton *P. vannamei* market shrimp

Assumptions: pond survival 75%; size at harvest 15g; FCR 1.2-1.5

| Group | PL production | | GO Production | | Total |
|------------------|---------------|------------|---------------|------------|---------------|
| | USD | % | USD | % | USD |
| feeds | 550 | 27.4 | 24,413 | 55.6 | 24,962 |
| labor | 536 | 26.7 | 5,167 | 11.8 | 5,702 |
| utilities | 686 | 34.2 | 7,750 | 17.6 | 8,436 |
| other | 81 | 4.0 | 930 | 2.1 | 1,011 |
| chemicals | 8 | 0.4 | 5,167 | 11.8 | 5,175 |
| health | 143 | 7.1 | 517 | 1.2 | 660 |
| sub total | 2,004 | 100 | 43,943 | 100 | 45,946 |
| management | 306 | | 5,167 | | 5,473 |
| capex | 284 | | 10,333 | | 10,617 |
| TOTAL | 2,594 | 100 | 59,443 | | 62,036 |

Not effective to drastically cut costs in hatchery phase

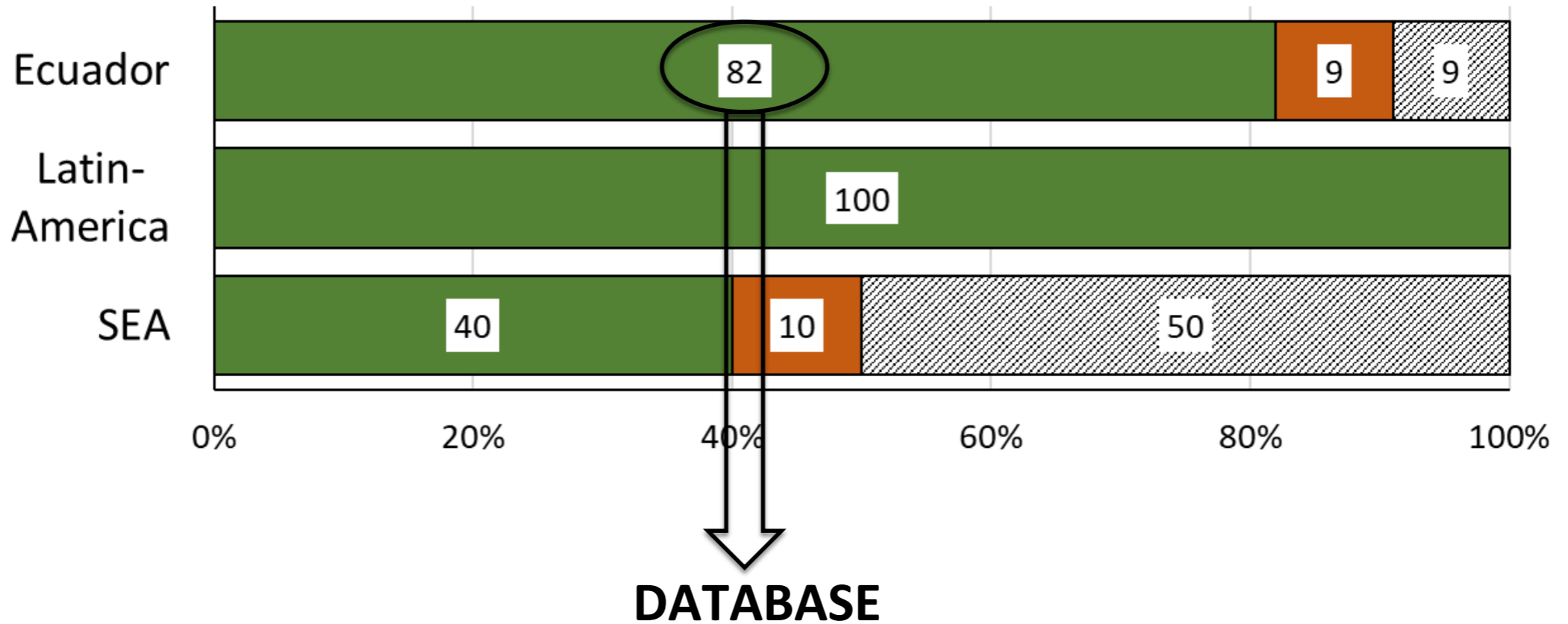
x 20

Gross margin is made in grow-out

4. Future

■ yes ■ no ■ blank

If you are convinced of a prolonged beneficial effect, do you have results that confirm this finding?



Unravel long-term, beneficial effects of **hatchery protocols** on **grow-out performance** through increased **PL quality**

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**THANK YOU
FOR YOUR ATTENTION!**

QUESTIONS?